

# IMPROVING THE EFFECTIVENESS OF CRUDE-OIL HYDROCARBON BIODEGRADATION EMPLOYING *AZOTOBACTER CHROOCOCCUM* AS CO-INOCULANT

Pujawati Suryatmana<sup>1</sup>, Edwan Karden<sup>2</sup>, Enny Ratnaningsih<sup>3</sup> and Wisjnuprpto<sup>2</sup>  
<sup>1</sup>Soil Science and Land Resource Department, Agriculture Faculty Padjadjaran University,  
Jatinangor 21<sup>st</sup>, Bandung Indonesia

<sup>2</sup>Environmental Engineering Study Programe, Institut Technology of Bandung, Ganesha 10<sup>st</sup> Bandung

<sup>3</sup>Chemical Study Programe, Institut Technology of Bandung, Ganesha 10<sup>st</sup> Bandung

## ABSTRACT

*Azotobacter chroococcum* has a great potential as *biosurfactant producing bacteria* and was used as co-inoculant to promote the rate of hydrocarbon biodegradation. The rate of hydrocarbon biodegradation were 0.01212, 0.01582 and 0.01766 g/l per day for *Acinetobacter* sp, *Bacillus cereus* and the consorsium culture respectively. On the other hand, the rates of biodegradation using *Azotobacter* as co-inoculant were 0.1472, 0.01612 and 0.02709 g/l per day. *Azotobacter chroococcum* co-inoculant has the capability of increasing biodegradation efficiency of crude oil hydrocarbon. The biodegradation efficiency of petroleum hidrocarbon was increated by 13.4, 14.6 and 14.4 % within the petrobacter cultures.

**Key words :** *Azotobacter chroococcum*, Biosurfactant, Co-inoculant, Petroleum hydrocarbon, Petrobacter, rate of biodegradation

## INTRODUCTION

The technology for exploiting the ability og microorganism to degrade crude-oil waste is still considered as one of the most efficient, economical, and environmentally sound methods (Margesin & Schinner 2001). Crude-oil biodegradation often encounters constraints because of its low hydrocarbon biodegradation rate. The rate of biodegradation is restricted by the mass transfer rate from solid phase to liquid phase, while the mass transfer rate is correlated with solubility level of target

compound. The low solubility of hydrocarbon, which comprises crude oil, is a problem in degrading the crude oil. Crude oil is always in the size of macrodroplets which can hamper the process of substrate sorption by oil-degrading bacteria (Van Eyk 1997).

Biosurfactant or bioemulsifier compounds can increase the solubility of crude-oil hydrocarbon.

5 *Rhodococcus rhodochrous* produces the extracellular compound S-2 (S-2 EPS), i.e. glucose, D-galactose, D-mannose, D-glucuronic acid, and fat. This compound can improve the degradation of aromatic fractions (AF) in crude oil. Beside functioning as a bioemulsifier for AF, the EPS compound can functioned as the carbon source for indigenous crude-oil degrading bacteria that to indigenous bacteria increase the growth rate of

10 One group of the microorganisms with an extracellular membrane that can act as bioemulsifier is *Azotobacter*. *Azotobacter* are nitrogen-fixing, non symbiotic bacteria. For example, *A. vinelandii* MTCC-2459 can produce an exopolysaccharide compound (EPS) that consists of glucose, rhaminose, galactose, and fucose (Vermani *et al.* 1997), while *A. chroococcum* produces extracellular product that can function as bioemulsifier (Wisjnuprpto *et al.* 2005). With the potential of this genus, *A.*  
15 *chroococcum* can be utilized as co-inoculant in biodegradation process of crude-oil hydrocarbon. *Azotobacter* are non-petrophylic bacteria (not hydrocarbon-degrading bacteria), but they can produce biosurfactant which in turn can increase the solubility of crude oil.

The objective of this research was to improve the rate and efficiency of crude-oil hydrocarbon biodegradation employing *A. chroococcum* as a co-inoculant.

20

## MATERIALS AND METHODS

***Azotobacter chroococcum* and Petrobacter.** The strain *A. chroococcum* used in this research was obtained from Soil Biology and Biotechnology laboratory, Faculty of Agriculture, Pajajaran University, Bandung. Petrobacter used in this research consisted of *Acinetobacter sp.* and *Bacillus*

*cereus*, while Petrobacter consortium consisted of *Acinetobacter* sp., *Bacillus cereus*, *Enterobacter* sp., and *Pseudomonas* sp. This petrobacter culture was isolated from an area polluted with textile industrial waste in Rancaekek, Bandung, and was known to have the ability of degrading hydrocarbon compounds (Data not shown).

5            Production media for *A. chroococcum* and Petrobacter was mineral media ( $K_2HPO_4$  1.5 g ,  $KH_2PO_4$  0.5 g ,  $MgSO_4$  0.5 g in 1.0 l aquadest) with glucose as carbon source, at pH 7. Media was sterilized at 121°C for 15 minutes. The fermentation for biosurfactant production was conducted at 28°C for three days with 100 rpm shaking speed. Biosurfactant used in this research was in the form of a three-day-old *A.chroococcum* culture, while for the Petrobacter the culture was 36 hours old.

10           To examine the biodegradation of crude-oil hydrocarbons, we used mineral media assay that contained crude oil as carbon source. Crude oil was obtained from Duri-Riau Field, Pertamina Balongan, Indramayu. The media was sterilized at 121°C for 15 minutes.

**Biodegradation of crude-oil hydrocarbons.** Bacterium *A.chroococcum* was grown in mineral media at 28°C for three days with 100 rpm shaking speed. This culture was used as biosurfactant for  
15   the degradation of crude-oil hydrocarbons conducted in a 1.0 litre closed culture. The medium was mineral media with 1 % (v/v) crude oil added as substrate. There were three kinds of Petrobacter cultures used in this research, i.e. *Acinetobacter* sp., *B. cereus*, and Petrobacter consortium consisted of *Acinetobacter* sp., *Bacillus cereus*, *Enterobacter* sp., and *Pseudomonas* sp.

This experiment was designed to be conducted in Batch reactor. There were three treatments of  
20   Petrobacter culture and another three treatments of Petrobacter culture plus *A. chroococcum* as co-culture. Overall, there were six combinations of treatments each with three replications. The density of the inoculant Petrobacter or *A. chroococum* used in every closed culture was 2% (v/v). Biodegradation process took place for 60 days. For the first 7 days samples were taken at 24-hour intervals. Afterwards, the interval was 7 days. The pH of the culture condition was adjusted to 7 and then

incubated at a 100 rpm shaking speed. Variables observed during biodegradation process were: cell viability, the reduction of total petroleum hydrocarbons (TPH) and residual compounds of biodegradation products. Closed cultures without *Petrobacter* or *A. chroococcum* were prepared as controls.

5           **Total Petroleum Hydrocarbons and Determination of its residual compounds.** The n-hexane extraction method of Iwabuchi (Iwabuchi *et al.* 2002, modified) was a modified and employed to isolate the residue of Total Petroleum Hydrocarbons from test cultures. Samplings for analysis were taken at 24-hour interval for the first 7 days and there after at 7- day interval.

10           The Gravimetric method was employed in TPH biodegradation. First stage TPH was extracted using n-hexane solvent. The ratio of sample to n-hexane solvent was 1:1, then the solution was then mixed using a vortex mixer at 10 rpm. The second stage of the process was separating TPH by centrifugation at 3500 g for 10 minutes. This stage produced three layers, at the bottom was the bacterial cells, in the middle was media residue, and at the top was crude oil dissolved in n-hexane. Oil and n-hexane were then poured into a bottle and the extraction process was repeated again until the  
15           solution became clear. The third stage was to recover the dissolved TPH through evaporation at 70°C. After that the weight of the TPH residue was determined.

20           After crude-oil degradation, hydrocarbon residues were analyzed using GC/MS. Analyses to check the types of compounds contained in the samples before and after processing were conducted at three observation points, i.e. at the beginning of the process, 14 days after starting the process, and at the end of the process. (GC separate the compounds in a gas phase by passing the gas flow through a stationary phase). Retention time in GC indicates the identity of the compounds . MS is a combination of mass analysis used to separate and identify the ions that comprise the components in a complex mixture.

## RESULTS

**Crude-oil Hydrocarbons and the Growth of Tested Bacteria.** The biodegradation process

5 by three *Petrobacter* cultures declined during the exponential growth phase of bacterial cells (Fig. 1). The rate of Crude-oil degradation by *B. cereus* decreased sharply during the first 14 days of incubation (Fig. 1a), while with the *Acinetobacter* sp., the decrease happened during the first 7 days (Fig. 1b). The reduction of Total Petroleum Hydrocarbons by *Petrobacter* consortium happened more quickly, also in the first 7 days when the growth was at exponential phase (Fig. 1c). After the exponential growth  
10 phase, the bacterial population declined, and biodegradation process continued slowly until the end of the incubation period.

**Specific Biodegradation Rate and Its Efficiency.** The specific TPH biodegradation rate by *B.*

*cereus*, *Acinetobacter* sp. and *Petrobacter* consortium increased with the addition of *A. chroococcum*  
15 (Table 1). The same also applied to the efficiency of crude-oil hydrocarbon biodegradation by *B. cereus*, *Acinetobacter* sp. and *Petrobacter* consortium which was higher compared to control, either with or without the addition of *A. chroococcum* (Table 2). The degradation rates were determined from the linear portion of the degradation curves.

20 **The number of the hydrocarbon compounds residues after biodegradation process.** The result of the analysis of hydrocarbon residual compounds qualitatively gave proven that *A. chroococcum* was capable of increasing the removal of TPH. It can be shown that the number of the TPH substances detected after biodegradation were qualitatively decreasing (the number of the hydrocarbon substances showed by the number of the peak area in Fig 2,3 and 4). The result showed

that the crude-oil components comprise 52 hydrocarbon compounds (t-0) dominated by aliphatic hydrocarbons, i.e. around 90% alkane compounds and 10% aromatic compounds such as benzene and naphthalene. Degradation by *B. cereus* during the first 14 days still left 37 hydrocarbon compounds as residue (Fig. 2a). The culture with *A. chroococcum* as co-inoculant had more effective degradation process; this was shown by 16 hydrocarbon compounds as residue (Fig. 2b). The residue from hydrocarbon degradation by *Acinetobacter* sp. still contained 42 compounds, while the addition of co-inoculant made the process higher with only 15 compound detected as residue (Fig. 3a, b). Likewise to hydrocarbon biodegradation by consortium culture, which was higher with the addition of coinoculant *A. chroococcu*. The residue from hydrocarbon degradation by Consortium. still contained 19 compounds, while the addition of co-inoculant made the process higher with only 15 compound detected as residue (Fig 4 a, b). That qualitatively data suggest that the removal of the crude oil hydrocarbon was enhanced in the petrobacter culture with the addition of *A. chroococcum*.

## DISCUSSION

In general, the bacteria used in this experiment actively utilized the substrate during logarithmic phase of their growth. This happened because crude oil component as the available hydrocarbons were in the form of aliphatic hydrocarbon groups that could easily be degraded and immediately be used as substrate by the bacteria. The bacterial growth after the exponential phase fluctuated (Fig. 1). This caused of the hydrocarbons were transformed into intermediate compounds by Petrobacter. These compounds could be toxic and disturb the growth and viability of Petrobacter's cells. The decline of viability could be interpreted as adaptation phase towards new substrate condition. Other than the presence of intermediate compounds, fluctuations happened because of substrate (carbon source) unavailability, unlike at the beginning of exponential phase when the substrate could be used immediately. Residual compounds as the result of transformation were often

resistant to biodegradation process. Besides, aggregation between the available hydrocarbon residual compounds frequently took place. Those compounds were difficult for the bacteria to utilize. This condition, that restricts continued degradation, also correlated with enzymatic activity rate that played a role in further catabolism process of intermediate compounds (Van Eyk 1997).

5           Bacteria needed time to adapt to, and deal, with the new substrate condition and resistant hydrocarbon residual compounds. Limiting factors in hydrocarbon degradation, among others, were low hydrocarbon solubility and slow hydrocarbon transformation from solid phase to liquid phase. Biosurfactant produced by *A. chroococcum* could increase substrate and hydrocarbon preparation to be immediately used by microorganisms. The enhancement of the TPH degradation rate was closely  
10   connected with the bacteria's uptake capacity to utilize the substrate. The higher uptake capacity the bacteria had, the more effective biodegradation process would be. Uptake process is closely related to mass transfer factor of the substrate in liquid phase. In the case of crude-oil hydrocarbons, mass transfer factor in liquid phase refers to solubility and dispersion level of oil droplets. Dispersion level is the level when large oil droplets can be emulsified and form oil micelles with maximum size  $< 1\mu\text{m}$   
15   which will make substrate absorption by *Petrobacter* easier.

Co-inoculant *A. chroococcum* could consistently improve biodegradation rate and the efficiency of crude-oil hydrocarbon biodegradation (Tabel 1 and 2). This bacterium had been confirmed acting as bioemulsifier that could increase the effectiveness of *Petrobacter* in the biodegradation process. This could happen because the extracellular substance produced by co-  
20   inoculant *A.chroococcum* could act as bioemulsifier. This bioemulsifier could improve dispersion and solubility of crude-oil hydrocarbons by forming oil micelles measuring  $0.5\mu\text{m}$ - $<1\mu\text{m}$ . These sizes of oil micelles were the suitable and desirable substrate size for *Petrobacter* cell to absorb them immediately so that hydrocarbon degradation process could be more effective. This was proven by the resistance and low solubility in hydrocarbon compound degradation process which happened after the

first week. The Hydrocarbon degradation process was improved with the presence of *A. chroococcum* as co-inoculant, it could increase the solubility of residual hydrocarbon compounds, which were difficult to dissolve and degrade. Co-inoculant *A. chroococcum*, a biosurfactant producing bacteria, could function as a bioemulsifier reservoir in an environment containing hydrocarbons. The biosurfactant mechanism in crude oil biodegradation is shown in Fig. 6. The *A. chroococcum* functioned as biosurfactant reservoir by forming oil micelles from large oil droplets. The outer layers of oil micelles were biosurfactant that had hydrophylic sequence, while the inner layer had hydrophobic sequence, these oil micelles acted as transport vehicles to form aggregates and performed crude-oil absorption process through bacterial cell wall (Van Eyk 1997). The bioemulsifier produced by *A. chroococcum* consisted of various fatty acid compounds acting as bioemulsifier that had amphiphatic characteristics (Suryatmana *et al.* 2006). These organic compound groups acted as Ligan Taxi System (transport vehicles) in the uptake process of hydrocarbons by *Petrobacter*. Therefore, *Petrobacter* could utilize the substrate more effectively.

The effect of biosurfactant on hydrocarbon degradation was based on increased hydrocarbon solubility and dispersion, and a change of the affinity between bacterial cell and hydrocarbon through induction that resulted in increased hydrophobicity of cell surface (Zhang & Miller 1995). With that mechanism, the rate and effectiveness of hydrocarbon adhesion was increased. The adhesion process was a very vital starting process in substrate uptake.

Consortium isolate showed the most effective characteristics in crude-oil hydrocarbon degradation. This was indicated by the highest biodegradation rate and efficiency. Likewise, *Petrobacter* consortium (more than one bacterial species) had higher probability of metabolic diversity compared to one bacterial species. In the consortium culture, combined performance in utilizing carbon source could happen between consortium members. A consortium of *Bacillus cereus*,



*Acinetobacter* sp., *Enterobacter* sp., and *Pseudomonas* sp. was a mix live that could positively interact and cooperate in using diverse substrate sources contained in crude oil.

Every *Petrobacter* isolate had a different level of ability in degrading aliphatic hydrocarbons contained in crude oil from Duri, like *B. cereus* and *Acinetobacter* sp. wich were able to degrade aliphatic hydrocarbon C7 to C18, while the consortium was able to degrade aliphatic hydrocarbons C7 to C22, (Fig 5). With those various metabolic abilities, the consortium culture prove more effective in TPH degradation activity compared to a single culture.

The result was indicated that intermediate compounds formed in the degradation process were in the form of various organic compounds dominated by fatty acids. The fatty acids might be an intermediate substances formed in alkane hydrocarbon degradation process by all the petrobacter used. The hypothetic of the alkane (docosane) biodegradation pathway by the consortium was showed in Fig 5. That hypothetic pathway was constructed based on the predominantly substances detected and was correlated by the aliphatic pathway of the Fritsche & Hofrichter's patern. The process from docosane (aliphatic hydrocarbon) may involved oxygenase enzyme systems. Alkane monoterminal oxidation pathway involves alcohol dehydrogenase formed docosanol. The alcohol dehydrogenase is intracellular enzyme which works together with electron carrier systems. Following aldehyde compounds (docosanal) formed from docosanol, that involve aldehyde dehydrogenase. The reaction process is followed by fatty acids (docosanoic acid) formation that requires  $\text{NAD}^+$ . The next process is where fatty acid assembled in cellular lipid and plays a role in provide energy through the  $\beta$ -oxidation pathway. The activation of fatty acid and coA results in the release of acetyl-coA, forming shorter fatty acids than alkane from original compounds, followed by the release of carbon dioxide. The acetyl coA then enters anabolic pathways to form new cells. In subterminal pathway reactions, alkane is changed into secondary alcohol before changed again into primary alcohol, and then transformed into fatty acid.

## ACKNOWLEDGMENTS

We thank Ministry of Research and Technology of Republic of Indonesia, for supporting this research through Riset Unggulan Terpadu XI (RUT XI) 2004 - 2006; Reginawanti Hindersyah, Dept. of Soil Science, Faculty of Agriculture, Padjadjaran University (UNPAD), for permission to use her isolate (*Azotobacter chroococcum*) in this research; Made Rhena Yasa (Head of Development Laboratory), PT PERTAMINA Indonesia, Balongan-Indramayu Production Unit, for the assistance given during this research.

## REFERENCES

Fritsche W & Hofrichter M. 2000. *Aerobic degradation by microorganisms*. In: Rehm HJ, G. Reed. (ed). *Biotechnology* Edisi ke 2. Weinheim: Wiley-VCH.

Iwabuchi N, Sunairi M, Urai M, Itoh C, Anzai H, & Harayama S. 2002. Extracellular polysaccharides of *Rhodococcus rhodochrous* S-2 stimulate the degradation of aromatic components in crude oil by indigenous marine bacteria. *Appl. Environ. Microbiol.* 68: 2337.

Margesin & Schinner. 2001. Bioremediation (natural attenuation and biostimulation) of diesel-oil-contaminated soil in an alpine glacier skiing area. *Appl. Environ. Microbiol.* 67: 3127-3133.

Suryatmana, P., Edwan K., Enny R., and Wisjnuprapto. 2006. Biosurfactant Characteristic of *Azotobacter chroococcum* . *J. Mikrobiologi Indonesia*. 11:30-34.

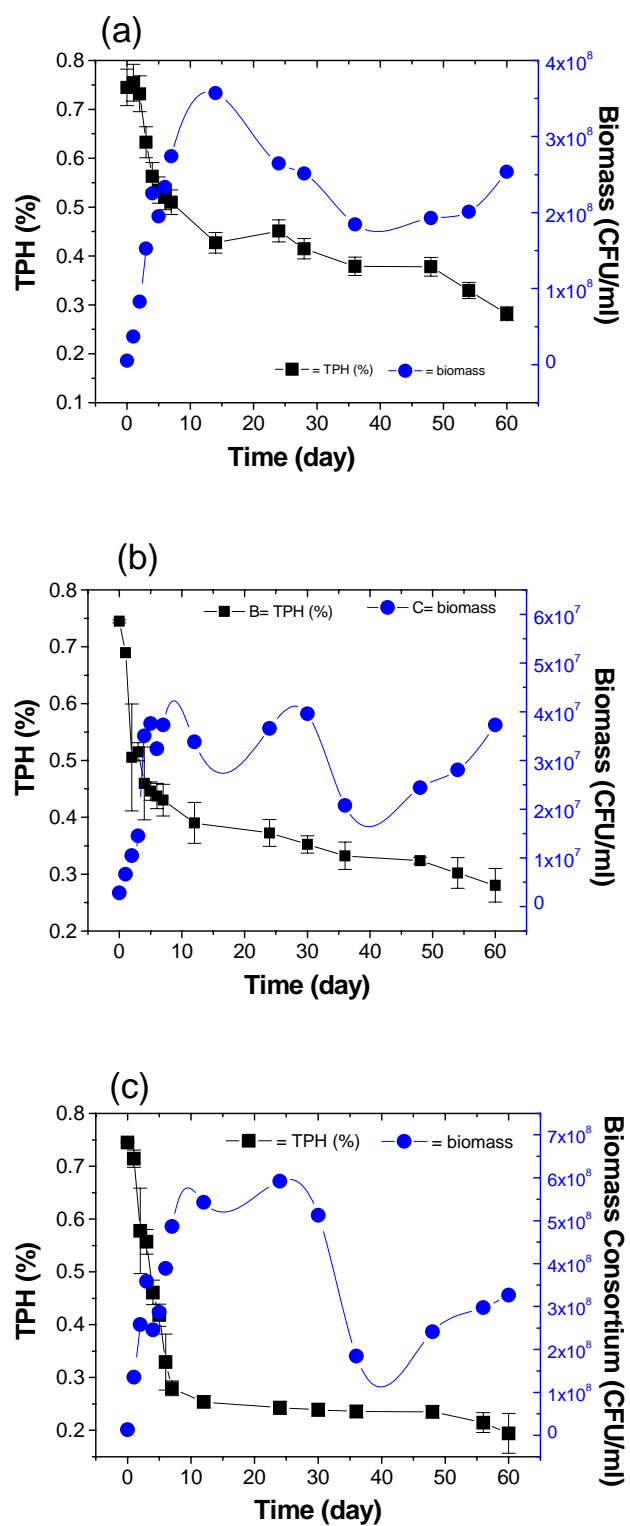
Van Eyk J. 1997. *Petroleum Bioventing*. A.A.Balkema. Rotterdam-Brookfield.

Vermani MV, Kelkar SM & Kramat MY. 1997. Studies in polysaccharide production and

growth of *Azotobacter vielandii* MTCC 2459, a plant rhizosphere isolate. *The Society for Applied Bacteriology, Letters in Applied Microbiology*. 24: 379 – 383.

5 Wisnuprpto., Edwan K., Pujawati., Gladys, S., Kristanti, N (2005). *Bioremediation of petroleum oil contaminated soils*. Proceedings of the COE Joint Symposium on Environmental Engineering Between Hokkaido University, Chungbuk National University and Bandung Institut of Technology. Sapporo-Japan, 2-4 February: 31-34.

Zang & Miller. 1995. Effect rhamnolipid (biosurfactant) structure on solubilization and biodegradation of n-alkanes. *Appl. Environ. Microbiol.* 61: 2247- 2251 .



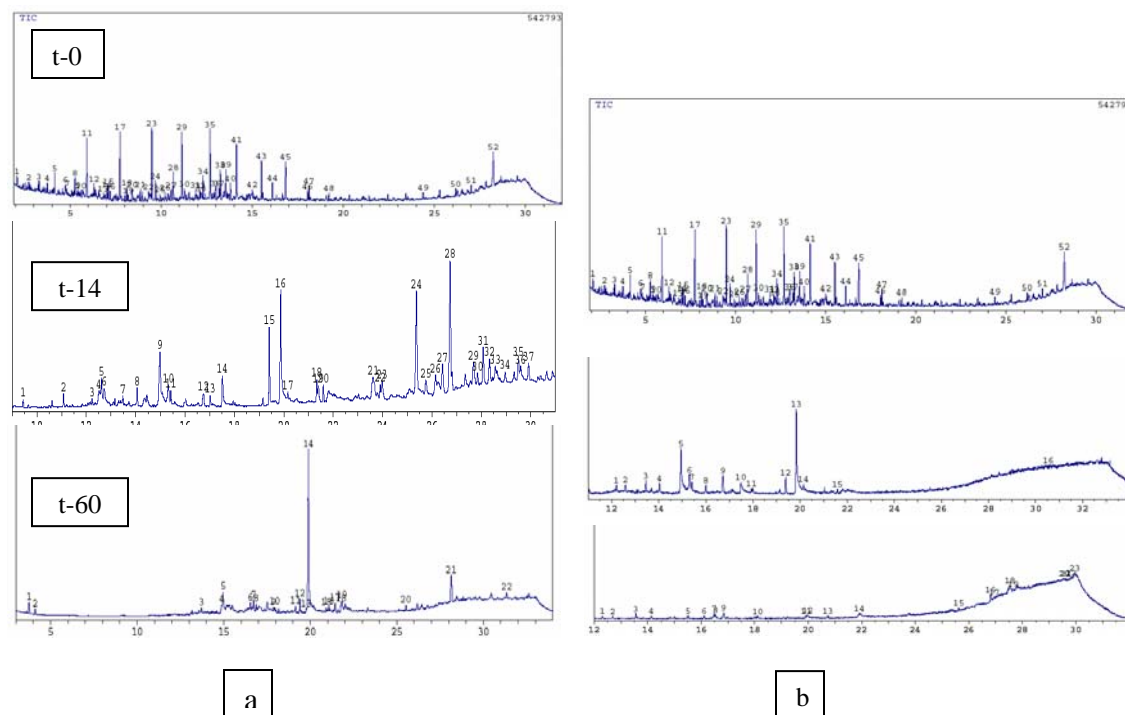
**Figure 1.** Total Petroleum Hydrocarbons and bacterial cell biomass during the growth of *Bacillus cereus* (a), *Acinetobacter* sp. (b), and *Petrobacter* consortium (c) in mineral media plus 1% crude oil over 60-day incubation period.

**Table 1.** Specific degradation rate of crude-oil hydrocarbons by three Petrobacter isolates.

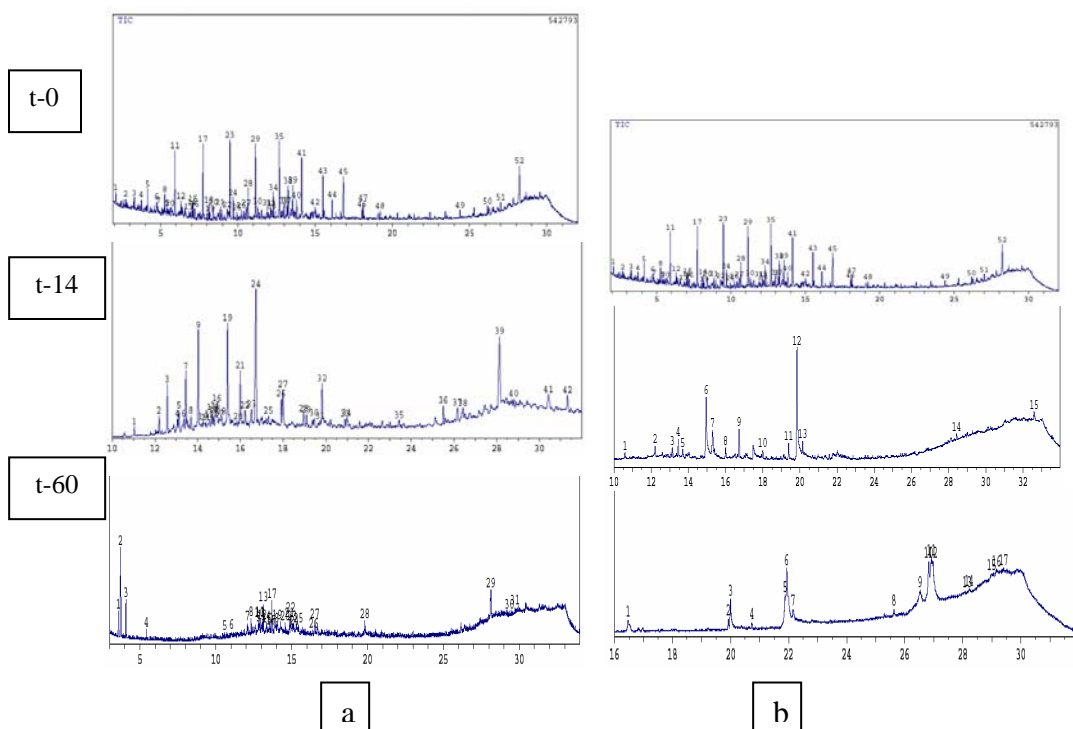
Isolate	Biodegradasi rate (g/l day <sup>-1</sup> )
<i>B. cereus</i> <i>B. cereus</i> + <b><i>A. chroococcum</i></b>	0.01582 <b>0.01612</b>
<i>Acinetobacter</i> sp <i>Acinetobacter</i> + <b><i>A. chroococcum</i></b>	0.01212 <b>0.01472</b>
Consortium Consortium+ <b><i>A. chroococcum</i></b>	0.01766 <b>0.02709</b>

**Table 2.** Efficiency of crude-oil hydrocarbon removal using Petrobacter treatment, with and without the addition of *A. chroococcum* (co-inoculant)

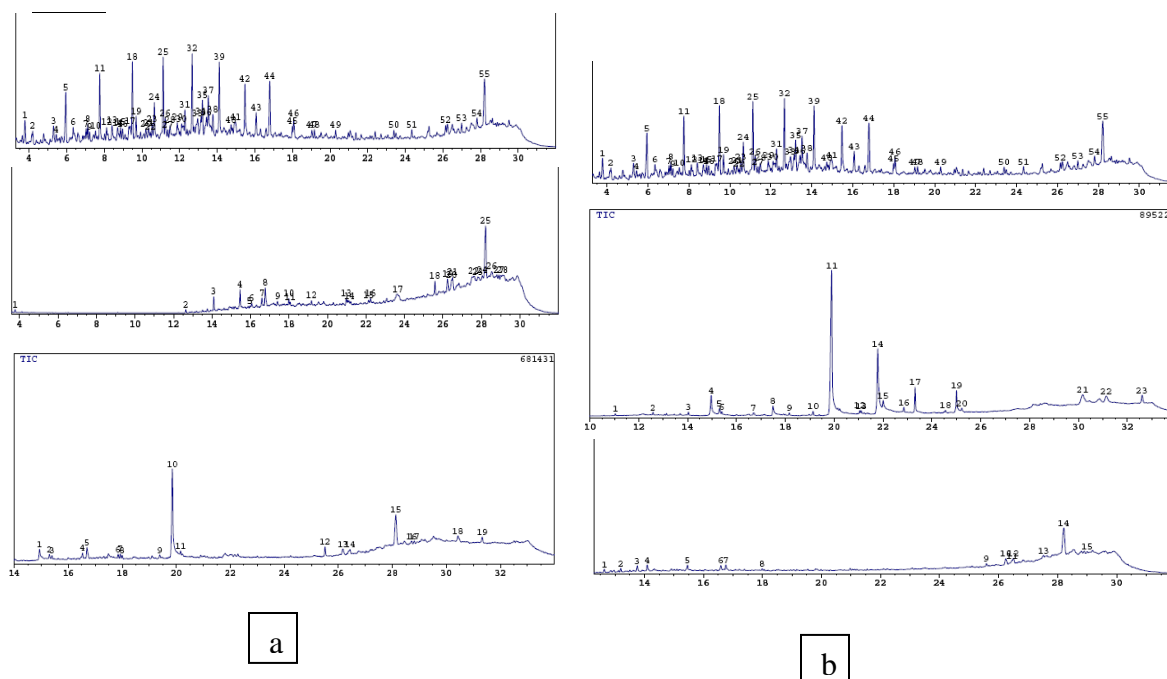
Petrobacter Isolate	TPH removal efficiency without <i>A. chroococcum</i> (%)	TPH removal efficiency with <i>A. chroococcum</i>	
		(%)	Enhancement TPH removal efficiency (%)
Control (no culture)	26.51 ± 1.33	39.67 ± 1.98	<b>13.16 ± 0.65</b>
<i>B. cereus</i>	62.17 ± 2.11	76.79 ± 1.84	<b>14.62 ± 0.27</b>
<i>Acinetobacter</i> sp.	57.26 ± 1.87	70.70 ± 1.53	<b>13.44 ± 0.34</b>
Consortium	71.18 ± 1.56	85.57 ± 2.28	<b>14.39 ± 0.07</b>



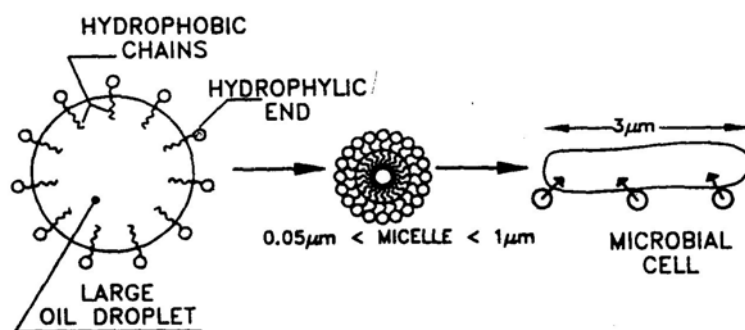
**Figure 2.** Chromatogram of crude-oil hydrocarbons before and after degradation by *B. cereus* (a) with the presence of co inoculant (b) on day 0, 14, and 60.



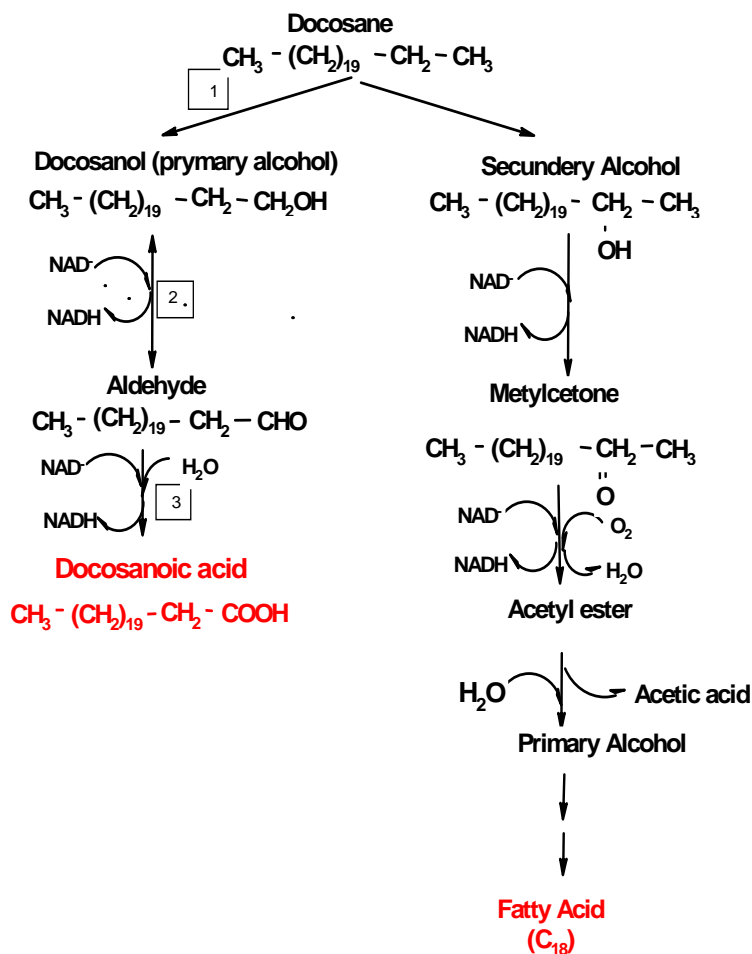
**Figure 3.** Chromatogram of crude-oil hydrocarbons before and after degradation by *Acinetobacter* sp. (a) with the presence of coinoculant (b) on day (t) 0, 14, and 60.



**Figure 4.** Chromatogram of crude-oil hydrocarbons before and after degradation by Consortium. (a) with the presence of co-inoculant (b) on day (t) 0, 14, and 60.



**Figure 6.** Model mechanism for crude-oil transportation and solubility by biosurfactant (Van Eyk, 1997)



**Figure 5.** The degradation pathway pattern of n-alkane: (1).The forming of docosanol (2), Aldehyde formed from primary alcohol; (3) Aldehyde was changed to the fatty acid (docosanoic acid).

*Note: The docosanoic acid and the fatty acids are the predominantly intermediate substances detected after 14 days biodegradation process*