

**ANTIMUTAGENIC ACTIVITY OF SINTOC BARK (*Cinnamomum sintoc*
Bl.)VOLATILE OIL WITH MICRONUCLEUS METHOD***

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Abstract

Sintoc bark (*Cinnamomum sintoc* Bl.) has many benefits, one of the efficacious compounds from these plants is volatile oil. Sintoc bark volatile oil are known to have antioxidant activity, antimutagenic test is carried out on rats using micronucleus method. The test materials were administered orally at a dose of 0.01 mL/200g, 0.025 mL/200 g, and 0.05 mL/200g. Antimutagenic effect evidenced by a decreasing number of micronucleus polychromatic erythrocytes per 1000 cells in preparations of bone smears. The results showed that volatile oil at all dose tested could reduce the number of micronucleus. Volatile oil dose 0.01 mL/200g decreased by 46.03%, 0.025 mL/200g dose of 38.73%, and a dose of 0.05 mL/200g of 29.21%. The final conclusion sintoc bark volatile oil a dose of 0.01 mL/200g antimutagenic effect best with the greatest reduction in micronucleus.

Key Word: Antimutagenic, *Cinnamomum sintoc* Bl.

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INTRODUCTION

Lifestyle and an unhealthy environment can cause a disease. On the environment and food products there are many compounds carcinogens (cancer-Causing agents) such as UV rays, industrial pollutants, pesticides, food additives, and products tobacco. These carcinogenic compounds can induce cancer by causing mutations that can change the arrangement of DNA. Mutations can be caused by the mutagenesis process of elimination, inversion, duplication, or transfer a particular DNA segment of the chromosome (Russell, 1994). One indicator of mutagenesis can be observed in living organism is micronucleus. Micronucleus is the result of mutations of chromosomes intact a broken / damaged and appears as a small nucleus within a cell. Micronucleus is easily observed in young cells polychromatic erythrocytes (reticulocytes). Number micronucleus in these cells reflect the level of genetic damage in a living system erythropoietic (Subarnas, et al., 2004). Micronucleus test is an in vivo screening procedure to detect any damage or loss of chromosomes (chromosome aberration) caused by mutagenic substances. Besides being used for testing mutagenic effects, micronucleus test can also be used for testing the effect of antimutagenic (Rhamdhani, 2003).

Usefulness of plants or herbs as antimutagenic and anticarcinogenic has been widely studied. Some of the natural constituents are believed to have antimutagenic or anticarcinogenic effects that potentially include: antioxidants, vitamins, essential oils, flavonoids, glucosinolat and organic sulfur compounds, tannins, polyphenols, and quinones . Volatile oil is one of the secondary metabolites produced by higher plants as the plants of the family Laminaceae, Lauraceae, Myrtaceae, and Rutaceae. *Cinnamomum sintoc* Bl. Essential oil are volatile at room temperature, containing more than one chemical compound and a typical smell. In the volatile oil contained compounds giving odor, fragrant, and typical of most essential oils are (Syafitri, 2006).

Sintok plants (*Cinnamomum sintoc* Bl.) is a type of plant with a woody stem that extends from the tribe and Lauraceae. Sintoc often used as an external drug or inside drugs . The part is often used as a drug is bark, bark of branches and leaves, while the fruit can be used as shampoo and soap. Sintok bark is used as an anthelmintic, the drug due to punctures and bites of poisonous animals, rheumatic (inflammatory), treatment of syphilis, to reduce intestine secreti (dysentery), and can eliminate the lower abdomen cramps (Heyne, 1987). In generally, toxicity test methods can be divided into two groups, first group consisted of toxicity test designed to evaluate the overall general effect of a compound in experimental animals, identified as an acute toxicity test, toxicity test subchronic, and chronic toxicity test. The

second class is specific toxicity test, the test potential, teratogenic test, and carcinogenesis tumorigenesis test, local effects on the skin of the eye, and behavioral test. In previous studies, toxicity test were conducted subchronic, and teratogenic test of sintoc bark. From the results of these studies, the bark does not give effect sintoc subchronic toxicity (Sumiwi, et al., 2008) and teratogenic effects (Subarnas, et al., 2008). To complement the pharmacological data and adds a sense of security from the use of sintok bark, then tested the carcinogenic or mutagenic in rat using micronucleus method. Based on the background the following problems can be identified whether sintok bark (*Cinnamomum sintoc* Bl.) volatile oil has antimutagenic activity.

This study aims to determine the antimutagenic activity of sintok bark (*Cinnamomum sintoc* Bl.) volatile oil as well as knowing the most effective antimutagenic doses of these volatile oil.

MATERIAL AND METHOD

Experimental animals

Experimental animals used were white male Wistar rats (200-250 g) \pm 2 months, obtained from PAU - ITB. Rats kept in the Laboratory of Pharmacology Faculty of Pharmacy, Unpad for approximately one month in cages with room temperature 23-28 ° C, humidity 70-85%, light settings 12 hours of light and 12 hours dark. Rats were fed pellets and drink water drinking bottles given unlimited (ad libitum).

Materials Chemistry

Antimutagenic activity test used siklofosfamida (N, N-bis (2-kloroetil) tetrahydro-2-1, 2,3, oksazafosforin-2-oxide) obtained from the pharmacy Galenika, Jakarta, and for Giemsa staining method used Merck Giemsa dye Solution obtained from Sakura Medical, Bandung.

Research Tools

Antimutagenic activity test used syringe 1 mL and 3 mL oral sonde, surgical equipment, sentrifugator Hettich type 2000 with a maximum rpm 6000 rpm, object glass, cover glass, light microscope brands Carton with up to 1000 times magnification, and the counter.

Method

Collection of Plant Material and Determination

Material in the form of sintok bark (*Cinnamomum sintoc* Bl.) obtained in Lembang, West Java. Bark is used as a form of sheets measuring ± 1 cm in order to obtain simplisia ready for further processing. Determinations carried out in the Department of Biology Faculty of Mathematics and Natural Sciences Padjadjaran University.

Isolation of Volatile Oil

Isolation of volatile Oils sintok bark in Installation Research Manoko Lembang, by steam distillation of water (water and steam distillation). Simplisia sintok bark incorporated into the distillation vessel, placed over boiling water using a bulkhead, so that plant material will only be fed with water vapor. The vessel is heated and distillation time is calculated from the first destilat dripping. Volatile oil distillation sintoc bark performed for 8 hours. Volatile oil obtained is collected in brown bottles to avoid direct contact with light.

Antimutagenic Assay

Antimutagenic activity assay of volatile oil bark sintoc performed using in vivo micronucleus method according to Schmid (Schmid, 1975) with modifications. Rats fasted approximately 18 hours before testing, but still provided enough drinking water. Rats were weighed, then were randomly divided into negative control group, positive control group and test group. Each group uses as many as 3 tails. Negative control group is the group of mice that received the suspension orally PGA 2% and 0.9% NaCl solution by intraperitoneal. Positive control group is the group of rats that received a solution of cyclophosphamide 50 mg / kg body weight intraperitoneally after administration of PGA suspension of 2% per orally. In the test group were given a solution of volatile oil prior to cyclophosphamide. Approximately 30 minutes after administration of volatile oil orally, every solution of the rats given cyclophosphamide 50 mg / kg intraperitoneally. After 24 hours, each rats readministered volatile oil and 30 minutes and then administered a solution of cyclophosphamide with the same dose as before. Six hours after the second

cyclophosphamide administration, the rats were killed and dissected to be taken both his thigh bones. Thigh bone is cleaned and marrow was taken by suction (aspiration). Marrow is inserted into a centrifuges tube containing physiological NaCl. Tube inserted into the sentrifugator at 1000 rpm for ten minutes. Most of the resulting supernatant removed with a pipette, the rest is mixed back with the resulting sediment. The liquid was then made preparations smear on objects glass and dried. Preparations allowed to stand for one day, then stained with Giemsa dye and covered with cover glass. Preparations were observed under a microscope magnification of 1000 times. For every 200 cells counted the number of micronucleus polychromatic erythrocytes contained in the polychromatic erythrocytes cells. The calculation is done as much as ten times in five different preparations for the rat tested. This means the number of polychromatic erythrocytes cells to be counted as many as 1000 cells to a mouse polychromatic erythrocytes. Criteria for counting cells that contain micronucleus polychromatic erythrocytes consist of: (1) micronucleus-shaped nucleus with a size of $1/20$ to $1/5$ the size of the nucleus. (2) Performed on cells that contain micronucleus polychromatic erythrocytes and not the amount of micronucleus in a cell. (3) The cell is calculated polychromatic erythrocytes are in intact (not broken or damaged). Antimutagenic activity of the test preparation is shown by a significant decrease in the number of micronucleus compared with the group.

Results

Collection fo materials and determination of plant

Determination sintoc bark showed that *Cinnamomum sintoc* Bl. included into the genus *Cinnamomum* (Lauraceae). Results determination can be seen in Image 1 and 2.



Figure 1. *Cinnamomum sintoc* Bl.

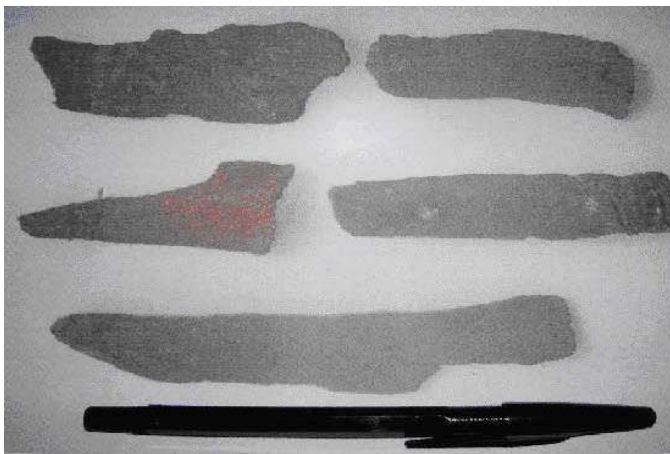


Figure 2. *Cinnamomum sintoc* Bl. bark

Volatile Oil Isolation

Results of sintok bark volatile oil isolation can be seen in Table 1.

Table 1. Results of Sintoc Bark Volatile Oil Isolation

| | |
|---------------------------|-----------|
| Weight Simplisia | 1000 gram |
| Volume volatile Oil | 5 mL |
| The Yield of volatile Oil | 0,5% v/w |

Antimutagenic Assay

Data obtained from the calculation of the amount contained micronucleus in 1000 polychromatic erythrocytes cells, where the observations are divided into five repetitions, and each time the loop consists of 200 cells polychromatic erythrocytes. Observations can be seen in Table 2.

Table 2. Number of Micronucleus 1000 Erythrocytes Polychromatic Cells

| Treatment | Duplication | | | Total | Everage | SD |
|------------------------------|-------------|-----|-----|-------|---------|-------|
| | 1 | 2 | 3 | | | |
| Normal Group | 22 | 16 | 30 | 68 | 22,67 | 19,90 |
| Negative Group | 306 | 323 | 316 | 945 | 315 | 8,54 |
| Volatile Oil 0,01mL/200g | 168 | 172 | 170 | 510 | 170 | 2 |
| Volatile Oil 0,025mL/200g | 192 | 199 | 188 | 579 | 193 | 5,57 |
| Volatile Oil 0,05 mL/200g | 230 | 224 | 215 | 669 | 223 | 7,55 |
| Total | | | | 2271 | | |

Number of micronucleus polychromatic erythrocytes per 1000 cells for all Groups treatment can be seen in Figure 3.

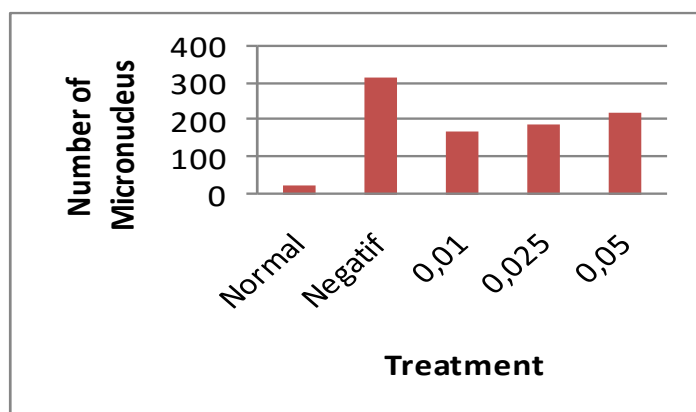


Figure 3. Bar graph of micronucleus polychromatic erythrocytes per 1000 cells for all treatment groups

Micronucleus polychromatic erythrocyte can be seen in Figure 3.

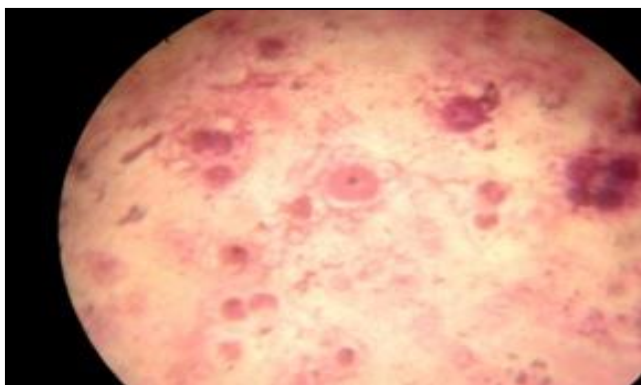


Figure 3. Micronucleus polychromatic erythrocyte can be seen in Figure 3.

Table 2 shows that in each treatment group there are differences in the number of micronucleus polychromatic erythrocytes per 1000 cells. An increasing number of micronucleus occurs in the negative control group due to administration of cyclophosphamide at a dose of 50 mg / kg compared with normal controls. This shows the effect of mutagenesis induced by cyclophosphamide induction. Cyclophosphamide which has been known as a chemotherapeutic agent for tumor disease can turn into a mutagen in the presence of metabolism in vivo. It has been known that cyclophosphamide need to go through the activation of enzymatic and nonenzymatis to produce its active form in the form of phosphoramidate mustard. Cyclophosphamide first hydroxylated to 4-hydroxycyclophosphamide through enzymatic hydroxylation process. The compound 4-hydroxycyclophosphamide in a state of equilibrium with tautomernya, namely aldophosphamide. 4- hydroxycyclophosphamide and aldophosphamide can undergo further metabolism in two lines, namely: changes to be non-toxic metabolites, such as 4-ketocyclophosphamide, carboxyphosphamide, and alcophosphamid; and changes into cytotoxic metabolites, namely acrolein and phosphoramidate mustard.

4-hydroxycyclophosphamide, acrolein and phosphoramidate mustard, is a alkylation compound a role in the toxic properties of cyclophosphamide. The damage produced by the reaction of alkylation compound begins with the occurrence of a covalent bond with a number of functional groups present on the cell components, such as-SH,-COOH,-NH and the phosphate groups of primary, hydroxyl and amino groups of nucleic acid bases. But the

position of the main bonding of the active metabolites of cyclophosphamide occurred in the DNA, which can occur in the process of crossing over that can cause DNA damage.

From Table 2 also can be seen a decrease in the number of micronucleus polychromatic erythrocytes per thousand cells are given volatile oil when compared with negative control. Number of micronucleus in the negative control decreased when given doses of volatile oil with 0.01, 0.025; 0.05 which was originally 315 to 170, 193, and 223. Based on statistical calculations using complete randomized design with 95% degree of confidence or $\alpha = 0.05$ in Appendix 2, is obtained that the calculated value of f greater than f_{α} tables. It shows that H_0 is rejected or there is the influence of the treatment group to decrease the number of micronucleus. Newman-Keuls test results showed any significant treatment where a single treatment with other treatments give different results. Based on these results can be demonstrated that administration of essential oil with a variety of doses have antimutagenic activity

Conclusion

Antimutagenic assay of sintoc bark volatile oil with variation dose using micronucleus method showed antimutagenic activity. This is shown by the decreasing number of micronucleus from 1000 polychromatic erythrocytes cells compared with negative control given cyclophosphamide 50 mg/bw. volatile oils 0.01 mL/200 g, 0.025 mL/200 g, and 0.05 mL/200 g decreased by 46.03%, 38.73%, and 28.21%. If viewed from the decrease in the number of erythrocyte micronucleus per 1000 cells, the volatile oil at a dose of 0.01 mL/200 g given the best antimutagenic effect with a decrease of 46.03%

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