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*Microbrewery*



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[Previous Issue](#)

*Beer is liquid bread*

Folk saying



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[This issue colophon](#)

20 articles from about 75 different authors

100 pages (2272 - 2371)

11 Microbiology articles

6 Food Sciences articles

3 Biotechnology articles

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#### ARTICLES FILTER

Show category:

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Microbiology

#### OPTIMIZATION AND EVALUATION OF MICROBE FORTIFIED COMPOSTS AS BIOCONTROL AGENTS AGAINST PHYTOPATHOGENIC FUNGI

Ajinath S. Dukare, Radha Prasanna, Lata Nain, Anil Kumar Saxena

Regular Article of Microbiology

[NLN DTD xml](#) | [Copernicus xml](#)

On this issue from page 2272 to page 2276.

[ABSTRACT AND DETAILS](#)

[direct link to fulltext pdf](#)

#### PROTEOLYTIC AND FIBRINOLYTIC ACTIVITIES OF HALOPHILIC LACTIC ACID BACTERIA FROM TWO INDONESIAN FERMENTED FOODS

Asep A. Prihanto, Darius, Muhamad Firdaus

## Short Communication of Microbiology

NLM DTD xml | Copernicus xml

On this issue from page 2291 to page 2293.

[ABSTRACT AND DETAILS ▶](#) [direct link to fulltext pdf](#)

## CHARACTERIZATION OF THE PARTIALLY PURIFIED PLANTARCIN SR18 PRODUCED BY LACTOBACILLUS PLANTARUM SR18

Wagih El-Shouny, Amal Abo-Kamar, Suzan Ragy

## Regular Article of Microbiology

NLM DTD xml | Copernicus xml

On this issue from page 2301 to page 2305.

[ABSTRACT AND DETAILS ▶](#) [direct link to fulltext pdf](#)

## THE EFFECT OF GROWTH PARAMETERS ON THE ANTIBIOTIC ACTIVITY AND SPORULATION IN BACILLUS SPP. ISOLATED FROM SOIL

Alev Usta, Elif Demirkan

## Regular Article of Microbiology

NLM DTD xml | Copernicus xml

On this issue from page 2310 to page 2313.

[ABSTRACT AND DETAILS ▶](#) [direct link to fulltext pdf](#)

## TITLE OF MANUSCRIPT IMPORTANT GROUPS OF MICROORGANISMS IN RAW GOAT MILK AND FRESH GOAT CHEESES DETERMINED DURING LACTATION

Libor Kalhotka, Květoslava Šustová, Michaela Hůlová, Jitka Přichystalová

## Regular Article of Microbiology

NLM DTD xml | Copernicus xml

On this issue from page 2314 to page 2317.

[ABSTRACT AND DETAILS ▶](#) [direct link to fulltext pdf](#)

## CELL-SURFACE BINDING OF DEOXYNVALENOL TO LACTOBACILLUS PARACASEI SUBSP. TOLERANS ISOLATED FROM SOURDOUGH STARTER CULTURE

Yousef I. Hassan, Lloyd B. Bullerman

## Short Communication of Microbiology

NLM DTD xml | Copernicus xml

On this issue from page 2323 to page 2325.

[ABSTRACT AND DETAILS ▶](#) [direct link to fulltext pdf](#)

## POLYPHASIC IDENTIFICATION OF CLOSELY RELATED BACILLUS SUBTILIS AND BACILLUS AMYLOLIQUEFACIENS ISOLATED FROM DAIRY FARMS AND MILK POWDER

Marcela J. González, Fernanda Gorgoroso, Stella M. Reginensi, Jorge A. Olivera, Jorge Bermúdez

## Regular Article of Microbiology

NLM DTD xml | Copernicus xml

On this issue from page 2326 to page 2331.

[ABSTRACT AND DETAILS ▶](#) [direct link to fulltext pdf](#)

## SCREENING OF SELECTED OLEAGINOUS YEASTS FOR LIPID PRODUCTION FROM GLYCEROL AND SOME FACTORS WHICH AFFECT LIPID PRODUCTION BY YARROWIA LIPOLYTICA STRAINS

Salinee Sriwongchai, Prayad Pokethitiyook, Maleeya Kruatrachue, Paramjit K. Bajwa, Hung Lee

## Regular Article of Microbiology

NLM DTD xml | Copernicus xml

On this issue from page 2344 to page 2348.

[ABSTRACT AND DETAILS ▶](#) [direct link to fulltext pdf](#)

## MICROBIOLOGICAL EVALUATION OF ANTIBIOTIC RESIDUES IN MEAT, MILK AND EGGS

Abdul Jabbar, Sajjad-Ur-Rehman

## Regular Article of Microbiology

NLM DTD xml | Copernicus xml

On this issue from page 2349 to page 2354.

[ABSTRACT AND DETAILS ▶](#) [direct link to fulltext pdf](#)

### SPECTROSCOPIC ANALYSIS OF FIVE PHYLOGENETICALLY DISTANT FUNGI (DIVISION: ASCOMYCETE) FROM VELLAR ESTUARY, SOUTHEAST COAST OF INDIA – A PILOT STUDY

Jayachandran Subburaj, T. R. Barathkumar, Visruth Prem, Muthusamy Thangaraj, Jeganathan Sivasubramanian

Regular Article of Microbiology



NLM DTD xml | Copernicus xml

On this issue from page 2355 to page 2359.

[ABSTRACT AND DETAILS ▶](#)

[direct link to fulltext pdf](#)

### ASSESSMENT OF BITTER LEAF (VERNONIA AMYGDALINA) ON SOME SELECTED PATHOGENIC MICROORGANISMS FROM UNIVERSITY OF ILORIN TEACHING HOSPITAL

Musa Olusegun Arekemase, Ganiyu Pacy Oyeyiola, Kabir Ishola Balogun

Regular Article of Microbiology



NLM DTD xml | Copernicus xml

On this issue from page 2360 to page 2365.

[ABSTRACT AND DETAILS ▶](#)

[direct link to fulltext pdf](#)

Food Sciences

### INVESTIGATION OF MOISTURE SORPTION BEHAVIOR OF AN INDIAN SWEET 'SON-PAPDI

Suni Bajpai, Pradeep Tiwari

Regular Article of Food Sciences



NLM DTD xml | Copernicus xml

On this issue from page 2277 to page 2282.

[ABSTRACT AND DETAILS ▶](#)

[direct link to fulltext pdf](#)

### QUALITY COMPOSITION AND BIOLOGICAL SIGNIFICANCE OF THE BANGLADESHI AND CHINA GINGER (ZINGIBER OFFICINALE ROSC.)

Sudam Nandi, Md. Moshfekus Saleh-e-In, Md. Matiur Rahim, Md. Nurul Huda Bhuiyan, Nasim Sultana, Md. Aminul Ahsan, Shamim Ahmed, Shajahan Siraj, Md. Zamilur Rahman, Sudhangshu Kumar Roy

Regular Article of Food Sciences



NLM DTD xml | Copernicus xml

On this issue from page 2283 to page 2290.

[ABSTRACT AND DETAILS ▶](#)

[direct link to fulltext pdf](#)

### TECHNIQUES FOR POTABLE WATER TREATMENT USING APPROPRIATE LOW COST NATURAL MATERIALS IN THE TROPICS

Sila Onesmus Nzung'a, Kotut Kiplagat, Okemo Paul

Regular Article of Food Sciences



NLM DTD xml | Copernicus xml

On this issue from page 2294 to page 2300.

[ABSTRACT AND DETAILS ▶](#)

[direct link to fulltext pdf](#)

### EFFECT OF PLASMA ENERGY ON THE ANTIOXIDANT ACTIVITY, TOTAL POLYPHENOLS AND FUNGAL VIABILITY IN CHAMOMILE (MATRICARIA CHAMOMILLA) AND CINNAMON (CINNAMOMUM ZEYLANICUM)

Solís-Pacheco J.R., Villanueva-Tiburcio J.E., Peña-Eguiluz R., González-Reynoso O., Cabrera-Díaz E., González-Álvarez V., Aguilar-Uscanga, B.R.

Regular Article of Food Sciences



NLM DTD xml | Copernicus xml

On this issue from page 2318 to page 2322.

[ABSTRACT AND DETAILS ▶](#)

[direct link to fulltext pdf](#)

### STATISTICAL OPTIMIZATION OF MEDIUM COMPOSITION AND PROCESS VARIABLES FOR XYLITOL PRODUCTION FROM RICE STRAW HEMICELLULOSE HYDROLYSATE BY DEBARYOMYCES HANSENI VAR HANSENI

Ramesh S., Muthuvelayudham R., Rajesh Kannan R., Viruthagiri T.

Regular Article of Food Sciences



NLM DTD xml | Copernicus xml

On this issue from page 2332 to page 2339.

[ABSTRACT AND DETAILS](#) ▶

 [direct link to fulltext pdf](#)

**PARTIAL HYDROLYSIS OF PURPLE SWEET POTATO FLOUR BY AMYLASE FROM SACCHAROMYCOPSIS FIBULIGERA AND ITS APPLICATION FOR COMPOSITE BREADMAKING**

Agus Safari, Dian S. Kamara, Fransiska Silalahi, Muhammad Fadhilillah, Idar Kardi, Safri Ishmayana

Regular Article of Food Sciences



NLM DTD xml | Copernicus xml

On this issue from page 2340 to page 2343.

[ABSTRACT AND DETAILS](#) ▶

 [direct link to fulltext pdf](#)

Biotechnology

**THE DISTRIBUTION OF CADMIUM AND COPPER IN FISH ORGANS**

Bartłomiej Zyśk

Short Communication of Biotechnology



NLM DTD xml | Copernicus xml

On this issue from page 2306 to page 2309.

[ABSTRACT AND DETAILS](#) ▶

 [direct link to fulltext pdf](#)

**STUDY OF GENETIC VARIABILITY OF TRITICALE VARIETIES BY SSR MARKERS**

Jana Ondroušková, Tomáš Vyhnanek

Short Communication of Biotechnology



NLM DTD xml | Copernicus xml

On this issue from page 2366 to page 2368.

[ABSTRACT AND DETAILS](#) ▶

 [direct link to fulltext pdf](#)

Previous Issue

**HEMATOLOGICAL ASSESSMENT OF ALBINO RATS FED WITH PLEUROTUS OSTREATUS CULTIVATED ON TWO TROPICAL TREES' SAWDUST (PYCNANTHUS ANGOLENSIS AND SPONDIAS MOMBIN)**

Soji Fakoya

Short Communication of Biotechnology



NLM DTD xml | Copernicus xml

On this issue from page 2369 to page 2371.

[ABSTRACT AND DETAILS](#) ▶

 [direct link to fulltext pdf](#)

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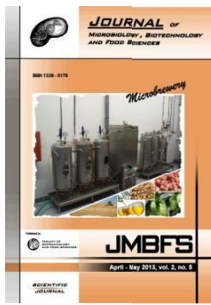
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## PARTIAL HYDROLYSIS OF PURPLE SWEET POTATO FLOUR BY AMYLASE FROM *Saccharomycopsis fibuligera* AND ITS APPLICATION FOR COMPOSITE BREADMAKING

Agus Safari, Dian S. Kamara, Fransiska Silalahi, Muhammad Fadhilillah, Idar Kardi, Safri Ishmayana\*

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### ABSTRACT

Purple sweet potato is one of underutilized carbohydrate sources in Indonesia, whilst known as good source of carbohydrate and can act as functional food due to its anthocyanine and dietary fiber contents. Therefore in the present study, we try to modify the sweet potato flour by partial hydrolysis using amylase produced by *Saccharomycopsis fibuligera* R64 and apply the partially hydrolyzed flour for composite breadmaking. The amylase was produced using batch method and partially purified by the addition of ammonium sulfate followed by gel filtration chromatography on Sephadex G25 using fast performance liquid chromatography system. The enzyme was then used to hydrolyze the purple sweet potato flour. Characterizations of the partially hydrolyzed flour comprise reduction in amylose-iodine complex, SEM and XRD. Partially hydrolyzed flour was then used as composite flour for bread, with ratio of wheat to partially hydrolyzed purple sweet potato flour was 70 : 30. The produced bread was then analyzed for its texture, organoleptic test and visualization of the bread crumb using TEM. The results of the present study indicate that the enzyme partially hydrolyzed the sweet potato flour. Even though the quality of the composite bread is not as good as wheat bread, partial hydrolysis seems to improve the texture and appearance of the composite bread, as indicated by better swelling volume and firmness of the composite bread using partially hydrolyzed purple sweet potato flour.

**Keywords:** Purple sweet potato, amylase, partial hydrolysis, composite bread

### INTRODUCTION

Purple sweet potato is one of carbohydrate source which can be considered as functional food due to its natural content of dietary fiber (Huang et al., 1999) and anthocyanin that can act as antioxidant, anticarcinogenic, antihypertensive and antimutagenic (Terahara et al., 1999; Oki et al., 2002; Kano et al., 2005). However, its application in various food products is still limited. Therefore there are many efforts has been conducted to develop various sweet potato based food product (Collado et al., 2001; Singh et al., 2004; Ahmed & Ramaswamy, 2006).

Bread is usually made from wheat flour because the presence of two specific proteins, gliadin and glutenin, that form protein network known as gluten which can seize carbon dioxide produced during fermentation of bread dough, which consequently make the bread swell (Goesaert et al., 2005). Unlike wheat, sweet potato does not have network forming protein. Instead, it has different type of protein, sporamin, that has been confirmed to have trypsin inhibitory activity (Shewry, 2003).

Enzyme application in breadmaking has been widely known to improve the quality of bread (Gerrard et al., 1998; León et al., 2002; Caballero et al., 2007). Amylases are one of the enzymes that are used to improve physicochemical properties of bread (Goesaert et al., 2009; Fadhilillah, 2011). However, as far as we aware, none of the published work used enzyme pretreated flour for making composite bread. Our lab has been working with amylase from locally isolated food-borne yeast (*Saccharomycopsis fibuligera* R64), which known to produce both  $\alpha$ -amylase and glucoamylase and it was also found that the  $\alpha$ -amylase belongs to mesophilic enzyme (Soemitro, 1996). The  $\alpha$ -amylase produced by this yeast was found to have raw starch degrading activity without adsorption mechanism, while the glucoamylase was found to adsorb onto starch granule (Hasan et al., 2008).

The objective of the present study was to investigate the effect of partial hydrolysis using partially purified amylase produced by *S. fibuligera* R64 and apply the partially hydrolyzed flour for composite bread making.

### MATERIAL AND METHODS

#### Production and partial purification of amylase

*S. fibuligera* cells were maintained on agar slant containing sucrose (6% w/v) and bacto agar (1.5% w/v) in bean sprout broth (10% w/v). Starter culture was developed by aseptically inoculated one yeast colony to 50 mL media containing sago starch (1% w/v) and yeast extract (1% w/v) for 48 hours at room temperature with 180 rpm shake speed. The starter culture was then transferred into a 500 mL fermentation media which has the same composition as starter culture media. The fermentation conducted for 72 hours at room temperature with 180 rpm shake speed. After 72 hours, the media was centrifuged at 4000×g to separate it from the cells. The supernatant contains enzyme which then partially purified by addition of ammonium sulfate (60-100% saturation), followed by gel filtration chromatography on Sephadex G-25 matrix (2 × 20 cm) using ÄKTAprime plus fast performance liquid chromatography system with 1 mL/min flow rate. The presence of protein and salt was detected using UV detector at 280 nm and conductivity meter attached to the system, respectively. Fractions with high absorbance at 280 nm were pooled and used for partial flour hydrolysis. Activity of the enzyme was monitored as described elsewhere (Hasan et al., 2008).

#### Partial hydrolysis of sweet potato flour

Purple sweet potato used in this study was *Ipomoea batatas* var. Ayamurasaki. The potato was peeled, washed and thin cut followed by sun dried. The dried chip was then grinded to obtain the sweet potato flour. Native purple sweet potato flour was referred as NF. Hydrolysis was conducted in 20% w/v suspension of sweet potato flour in 50 mM phosphate-citrate buffer pH 5.8. Partially purified enzyme was added to achieve 50 unit/g flour. The hydrolysis was performed at two different temperatures, i.e. the optimum temperature of enzyme activity (50°C) and room temperature for 12 hours. The flour was then separated by centrifugation at 4000×g and the reducing sugar content of the supernatant was determined using alkaline ferricyanide assay (Walker & Harmon, 1996). The resulting flour was referred as HT<sub>r</sub> and HT<sub>o</sub> for partially